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Effects of the 5-HT₇ receptor antagonist SB-258741 in animal models for schizophrenia

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Abstract

The 5-HT₇ receptor is targeted by several new antipsychotics such as clozapine and risperidone. We studied the effect of R-(+)-1-(toluene-3-sulfonyl)-2-[2-(4-methylpiperidin-1-yl)ethyl]-pyrrolidine (SB-258741), a specific 5-HT₇ receptor antagonist, in three models for positive symptoms, D-amphetamine-induced hyperactivity and D-amphetamine- and phencyclidine (PCP)-disrupted prepulse inhibition (PPI) in rats, with the aim of investigating the role of this receptor in the clinical effect of antipsychotics. We also tested this compound in a model for negative symptoms, PCP-disrupted social interaction (SIT) in rats. Induction of side effects by this compound was evaluated by testing its potency to reduce spontaneous motility and to induce catalepsy in rats. The effect of SB-258741 was compared to risperidone in all models. This study showed that SB-258741 had no beneficial effect on PCP-disrupted SIT. SB-258741 did not reverse D-amphetamine-disrupted PPI; however, it dose-dependently normalised PCP-disrupted PPI. SB-258741 antagonised D-amphetamine-induced hyperactivity but reduced motility of rats at similar doses. Thus, this specific 5-HT₇ receptor antagonist brought a clear positive outcome in only one model for positive symptoms of schizophrenia and had no beneficial effect in the model for negative symptoms. Consequently, it is clear that SB-258741 affects the PPI phenomenon but is not expected to have an antipsychotic effect on its own in clinic. © 2002 Elsevier Science Inc. All rights reserved.

Keywords: SB-258741; 5-HT7; Prepulse inhibition; Schizophrenia; Activity; Social interaction; Catalepsy

1. Introduction

The 5-HT₇ receptor is targeted by several antipsychotics of second generation such as clozapine, risperidone, and zotepine, and with lower affinity, by olanzapine and sertindole (Arnt and Skarsfeldt, 1998; Eglen et al., 1997; Roth et al., 1994; Terrón and Falcón-Neri, 1999). In in vitro assays, clozapine also up-regulates the expression of the 5-HT₇ receptor (Zhukovskaya and Neumaier, 2000). The localisation of the 5-HT₇ receptor at the level of limbic structures (Branchek et al., 1995; Doyland et al., 1995; Hagan et al., 2000; Le Corre et al., 1997; Thomas et al., 1999; To et al., 1995) also suggests a possible role for treatment of schizophrenia. However, the role per se of this receptor in the clinical action of antipsychotics is still unknown. Various in vivo studies have been conducted in order to define the role of this receptor, and sometimes with nonspecific compounds (Meneses and Terrón, 2001). This

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receptor has been suggested to be of interest for treatment of cognitive dysfunction (Meneses and Terrón, 2001), depression (Yau et al., 1997; Schwartz, 1993; Sleight et al., 1995), and sleep disorders (Schwartz, 1993; Tsou et al., 1994) by regulation of circadian rhythms (Lena Mullins et al., 1999; Lovenberg et al., 1993). However, no preclinical studies described its role in animal models related to schizophrenia.

This lack of knowledge is basically due to a lack of potent and specific 5-HT₇ ligands. Recently, 5-HT₇ receptor antagonists have been developed (Forbes et al., 1998; Hagan et al., 2000; Lovell et al., 2000), R-(+)-1-(toluene-3-sulfonyl)-2-[2-(4-methylpiperidin-1-yl)ethyl]-pyrrolidine (SB-258741) being one of the most specific (see compound "13" in Lovell et al., 2000).

In order to clarify the potential role of a 5-HT₇ receptor antagonist for the treatment of schizophrenia, we studied the effect of SB-258741 in three models related to positive symptoms of schizophrenia, D-amphetamine-induced hyperactivity and D-amphetamine- and phencyclidine (PCP)-disrupted prepulse inhibition (PPI) in rats. We also tested this compound in a putative model for negative symptoms, PCPdisrupted social interaction (SIT) in rats. Induction of motor

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side effects by this compound was evaluated by testing its potency to reduce spontaneous motility and to induce catalepsy in rats.

The D-amphetamine-induced hyperactivity in rats has the advantage of being a predictive and reproducible model for the selection of compounds with antipsychotic-like action (Arnt, 1995, 2000). Consequently, it should indicate whether SB-258741 has a general antipsychotic potential.

PPI (Braff and Geyer, 1990; Swerdlow et al., 1994) is disrupted in patients with schizophrenia (Swerdlow et al., 1994; Braff and Grillon, 1992), and this is modeled in the rat by systemic administration of the indirect dopamine (DA) agonist D-amphetamine (Druhan et al., 1998; Swerdlow et al., 1986; Mansbach et al., 1988; Zhang et al., 2000; Johansson et al., 1995; Paabøl Andersen and Pouzet, 2001; but see Davis, 1988), or the noncompetitive N-methyl-Daspartate (NMDA) antagonist PCP (Mansbach and Geyer, 1989; Bakshi et al., 1994). PPI disruption in rodents is antagonised by antipsychotic drugs, and this effect is used as an animal model of antipsychotic action. Interestingly, while disruption of PPI by a dopamine enhancer is reversed by both classical and the second generation of antipsychotics (Swerdlow and Geyer, 1993; Swerdlow et al., 1994; Yamada et al., 1999; Paabøl Andersen and Pouzet, 2001), PPI disruption produced by noncompetitive NMDA antagonists is apparently specifically reversed by the second generation of antipsychotics (Bakshi and Geyer, 1995; Bakshi et al., 1994; Yamada et al., 1999). On the other hand, a lack of effect in this model has also been claimed for clozapine or risperidone (Johansson et al., 1994; Swerdlow et al., 1996). Thus, we investigated the effect of SB-258741 in D-amphetamine-disrupted PPI in order to determine the possible general antipsychotic-like effect of this compound, and we considered of interest to test the effect of SB-258741 in PCP-disrupted PPI in order to define similarities with the new generation of antipsychotics.

A specific advantage of the second generation of antipsychotics is that they have a lower potential to induce extrapyramidal side effects (EPS) than classical antipsychotics (Goldstein, 2000; Glazer, 2000), and they also partially reduce negative symptoms such as social withdrawal in schizophrenic patients (Blind, 1999). In order to illustrate the capacity of compounds to reduce social withdrawal, we studied their effect in PCP-disrupted SIT in rats (Corbett et al., 1995; Sams-Dodd, 1995). Few antipsychotics (clozapine, risperidone, and sertindole) show capacity to antagonise PCP-disrupted SIT (Sams-Dodd, 1997; 1998). As these three antipsychotics (Arnt and Skarsfeldt, 1998; Roth et al., 1994) have affinity for the 5-HT₇ receptor, it was consequently of interest to test whether SB-258741 would have a positive effect in this model as well.

The effect of SB-258741 was compared to risperidone in all models tested, as this antipsychotic is the one on the market with the highest affinity for the 5-HT₇ receptor (Roth et al., 1994). The potential interest of a 5-HT₇ receptor antagonist for treatment of schizophrenia will be discussed.

2. Materials and methods

2.1. Subjects

Rats used in all experimental paradigms except PPI were male Wistar rats supplied by Møllegård (Denmark). Rats used in the PPI were supplied by Charles River (Germany). Animals were approximately 2 months old and weighed approximately 250-300 g when used in the PPI and SIT paradigm, but 200-250 g when used in the catalepsy, motility, and D-amphetamine hyperactivity paradigms. New animals were used in each paradigm. Animals were housed four per cage (macrolon type III) for the motility, D-amphetamine hyperactivity, and catalepsy paradigms, but two per cage for the SIT and PPI paradigms. They were kept in climate-controlled animal facilities (temperature at 21 ± 2 °C and relative humidity at $60 \pm 10\%$). Animals were housed under a normal light-dark cycle (lights on 06:00-18:00 h) with free access to food and water, except for the PCP-disrupted SIT model, in which animals were housed under a reversed light-dark cycle (lights on 18:00-06:00 h). Animals were randomly divided into treatment and drug groups. All procedures were in strict accordance with the Danish Committee on Care and Use of Laboratory Animals.

2.2. Materials and experimental design

2.2.1. D-Amphetamine-induced hyperactivity and motility

The test cages were macrolon type III, high model $(42.5 \times 26.5 \times 18.5 \text{ cm})$ equipped with four infrared light sources and photocells 4 cm above the bottom of the cages. The D-amphetamine-induced hyperactivity experiments were run in normal light conditions in an undisturbed room. The test substances were injected subcutaneously (sc) 30 min before injection (sc) of D-amphetamine sulphate (0.5 mg/kg). Immediately after injection of D-amphetamine, the rats were placed individually in test cages, and locomotor activity was measured in 15 min intervals for 2 h. The motility experiments were performed in identical cages, but were located in a dark room in order to obtain a better spontaneous activity and, consequently, optimal conditions to highlight possible motor inhibition after administration of test substances. The test substances were injected subcutaneously 30 min before measuring motility for 15 min, which means during the exploratory phase of rats.

2.2.2. Prepulse inhibition

The apparatus consisted of four startle chambers (MOPS 2b; Metod och Produkt, Göteborg, Sweden). Each rat was placed in a wire-mesh cage $(18.5 \times 7 \times 6.5 \text{ cm})$ that was suspended at one point to a piston within a stabilimeter in such a way that this cage could move freely under the piston. Each cage was enclosed within an individual sound-insulated box $(52 \times 42 \times 38 \text{ cm})$. Each movement of the piston was converted to an analogue signal by an accelerometer. Signals were transferred to a computer using an

analog-to-digital converter from National Instruments. Custom-designed StarWin software package from Ellegaard Systems (Denmark) ran the startle-stimuli schedules and the real-time data analysis. Acoustic noise bursts were presented via two speakers mounted 15 cm behind the cages. A background noise (62 or 70 dB) was maintained throughout the session. Startle amplitude was defined as the maximal of one hundred 1-ms stabilimeter readings collected from stimulus onset. The four cages were calibrated for equal sensitivity before starting experimental sessions. Stimulus consisted of a burst of white noise superimposed to the background noise with a rise time of less than 1 ms.

The PCP-disrupted PPI procedure was designed as follows. After a 10 min acclimatization with the background sound on, eight startle pulses of 105 dB broad band burst for 30 ms were presented to test for basal startle responsiveness. Then eight blocks of six different trial types were presented to measure PPI. Trials were presented pseudorandomly throughout the session, i.e., pulse alone (105 dB), prepulse alone (77 dB), prepulse followed by pulse (three trial types: 67+105 dB; 72+105 dB; 77+105 dB) or no pulse. The three different prepulses had an intensity of either 5, 10, or 15 dB above the background sound (62 dB) and a duration of 20 ms. The time interval between the prepulse offset and the pulse onset was 60 ms. The intertrial period was constant and lasted 15 s. The percentage PPI induced by each prepulse intensity was calculated as: $[100 - (100 \times \text{startle amplitude})]$ on prepulse trial)/(startle amplitude on pulse alone trial)]. PCP (2 mg/kg) or vehicle was injected subcutaneously 10 min before starting acclimatization. Compounds to be tested were injected 15 min before PCP administration.

The D-amphetamine-disrupted PPI design differed for some parameters. After a 5 min acclimatization period with the background sound on, eight startle pulses of 120-dB broad band burst for 40 ms were presented to test for basal startle responsiveness. The three different prepulses had an intensity of either 4, 8, or 12 dB above the background sound (70 dB) and a duration of 30 ms. The time interval between the prepulse offset and the pulse onset was 100 ms. The intertrial period was constant and lasted 15 s. D-Amphetamine (2 mg/kg) or vehicle was injected subcutaneously 25 min before starting acclimatization. Compounds to be tested were injected 15 min before D-amphetamine administration.

2.2.3. Social interaction

The test was performed in an open arena $(150 \times 100 \times 40 \text{ cm})$. The behaviour of rats was recorded by a video camera placed above the arena and connected to an S-VHS video recorder. Lighting in the room consisted of dark-red diffused light. Videos were analysed off-line by the Ethovision programme (Noldus) in order to evaluate activity, active SIT, and passive SIT of rats. SIT is defined as the duration during which animals were less than 20 cm from each other. SIT was subsequently divided into an active and passive component for each rat based upon whether the rat

moved or was inactive (Sams-Dodd, 1996). The method applied for this experiment is the same as the 3-day treatment protocol described in detail elsewhere (Sams-Dodd, 1997). To summarise shortly, 2 weeks after arrival in our facilities, half of the rats within a drug group were dyed with black hair colour except on the head. Rats received a daily injection of SB-258741 and PCP (2 mg/kg) for 3 consecutive days. Subjects were tested on the last day of injection. SIT of rats was measured for 10 min after placing two unfamiliar rats (one white and one dyed black) in the open arena, on the same wall side but at opposite corners. Both rats received the same drug treatment.

2.2.4. Catalepsy

Catalepsy was assessed on a vertical wire mesh frame $(50 \times 50 \text{ cm})$. Mesh opening was $1 \times 1 \text{ cm}$ and mesh diameter was 2 mm. Animals were considered cataleptic when they remained immobile during a period of 15 s. Rats showing muscle relaxation were not considered cataleptic. Rats that did not move their paws but showed active body or head movements were also not considered as cataleptic. Observation for catalepsy occurred once each hour during the first 6 h and once 24 h after dosing.

2.3. Drugs

PCP hydrochloride was synthesized at H. Lundbeck and dissolved in 0.1 M methanesulfonic acid diluted in 0.9% NaCl. D-Amphetamine sulfate was supplied by Nomeco (Copenhagen) and was dissolved in 0.9% NaCl. SB-258741 was synthesised at H. Lundbeck and was dissolved in water. Risperidone was a courtesy of Janssen Pharmaceutical (Beerse) and was dissolved in 0.1 M HCl diluted in 0.9% NaCl. Solution used as vehicle control was always 0.9% NaCl. All compounds were injected at volumes of 5 ml/kg. Except for PCP and D-amphetamine, all doses are expressed in milligrams per kilogram, according to the amount of free base.

2.4. Data analysis

2.4.1. D-Amphetamine-induced hyperactivity

Hyperactivity was analysed by a two-way repeated measurement ANOVA consisting of a between-subjects factor of treatment (six levels for SB-258741: VEH– VEH; VEH–AMPH; 0.56 mg/kg AMPH; 2.3 mg/kg AMPH; 4.6 mg/kg AMPH; 9.1 mg/kg AMPH; five levels for risperidone: VEH–VEH; VEH–AMPH; 0.080 mg/kg AMPH; 0.31 mg/kg AMPH; 1.3 mg/kg AMPH) and a within-subjects factor of Time (eight bins of 15 min each). A post hoc analysis Fisher's PLSD was performed to compare between groups and time intervals.

2.4.2. Motility

Motility was analysed by a one-way ANOVA consisting of a between-subjects factor of drug (five levels for SB- 258741: VEH; 1.1 mg/kg; 2.3 mg/kg; 4.6 mg/kg; 9.1 mg/kg; four levels for risperidone: VEH; 0.31 mg/kg; 1.3 mg/kg; 4.9 mg/kg). A post hoc analysis Fisher's PLSD was performed to compare between groups.

2.4.3. D-Amphetamine- or PCP-disrupted PPI

The startle amplitude was analysed by a two-way ANOVA consisting of a between-subjects factor of treatment (two levels: VEH; and AMPH or PCP) and a betweensubjects factor of drug (three levels: VEH; 2.3 mg/kg; 9.1 mg/kg, for SB-258741; and VEH; 0.080 or 0.16 mg/kg; and 0.16 or 0.31 mg/kg for risperidone). Mean percentage PPI was analysed by three-way repeated-measures ANOVA consisting of a between-subjects factor of treatment, a between-subjects factor of drug, and a within-subjects factor of three prepulse intensities. A post hoc analysis Fisher's PLSD was performed to compare the startle amplitude and the mean percent PPI between groups.

2.4.4. PCP-disrupted SIT

Activity, active SIT, and passive SIT were analysed by a two-way ANOVA consisting of a between-subjects factor of treatment (two levels: VEH and PCP) and a between-subjects factor of drug (three levels: VEH; 2.3 mg/kg; 9.1 mg/kg, for SB-258741; and VEH; 0.16 mg/kg; 0.31 mg/kg, for risperidone). A post hoc analysis Fisher's PLSD was performed to compare activity and SIT between groups.

2.4.5. Catalepsy

The total catalepsy score was expressed as percent of the maximum achievable score for the individual rat and the result presented as mean values per treatment group. Catalepsy was analysed by a two-way repeated-measures ANOVA consisting of a between-subjects factor of drug (two levels for SB-258741: 9.1 and 18 mg/kg; three levels for risperidone: 2.5 mg/kg; 5.0 mg/kg; 10 mg/kg), and a within-subjects factor of Time (seven levels: 1, 2, 3, 4, 5, 6, and 24 h).

All analyses were based on the raw data and were calculated with the SigmaStat (2.03) software system, except for the PPI data, which were analysed with the StatView (5.0) software system. Preliminary results of this study were presented as a poster at the SFN meeting 2000.

3. Results

3.1. D-Amphetamine-induced hyperactivity

3.1.1. SB-258741

D-Amphetamine-induced hyperactivity (Fig. 1) as shown by the significant difference of activity between the VEH– VEH- and VEH–AMPH-treated groups on the time intervals 30–105 min postinjection (*P*'s < .01). The post hoc analysis conducted on the significant Treatment × Time interaction, F(35,406)=7.40, P<.001, showed that SB-



Fig. 1. Effects of acute treatment with SB-258741 on D-amphetamineinduced (0.5 mg/kg) hyperactivity in rats. n=8 per Drug × Treatment group, but n=16 in VEH–VEH and VEH–AMPH groups. *P<.05 versus VEH–VEH; #P<.05 versus VEH–AMPH.

258741 administered at 0.56 mg/kg did not reverse the effect of D-amphetamine. SB-258741 administered at 2.3 mg/kg reversed hyperactivity induced by D-amphetamine up to 45 min after starting measurement (P's < .04). SB-258741 given at the two highest doses (4.6 and 9.1 mg/kg) reversed hyperactivity induced by D-amphetamine up to 60 min after starting measurement (P's < .03 and P's < .002, respectively). At 15 min after starting the observation, the two highest doses of SB-258741 given in addition to D-amphetamine reduced motor activity below that of VEH–VEH-treated animals (P's < .001). On the time interval 120 min, the groups treated with the three highest doses were more active than the VEH–AMPH group (P's < .03) and the VEH–VEH group (P's < .003).

3.1.2. Risperidone

D-Amphetamine-induced hyperactivity (Fig. 2) as shown by the significant difference of activity between the VEH–VEH- and VEH–AMPH-treated groups on the time intervals 30–105 min postinjection (P's < .03). The post hoc analysis conducted on the significant Treatment × Time interaction, F(28,245) = 6.84, P < .001, showed that only risperidone given at the two highest doses (0.31 and 1.3 mg/kg) reversed hyperactivity induced by D-amphetamine up to 45 and 60 min after starting measurement (P's < .001). During the first 30 min after starting the observation, the highest dose of risperidone (1.3 mg/kg) given in addition to D-amphetamine reduced activity of rats below the level of VEH–VEH-treated animals (P's < .005).

3.2. Motility

3.2.1. SB-258741

A significant main effect of drug, F(4,27) = 7.59, P < .001, demonstrated that SB-258741 tested at 4.6 and 9.1 mg/kg reduced motility of rats (P's < .03) (Fig. 3).



Fig. 2. Effects of acute treatment with risperidone on D-amphetamineinduced (0.5 mg/kg) hyperactivity in rats. n=8 per Drug × Treatment group. *P<.05 versus VEH–VEH; #P<.05 versus VEH–AMPH.

3.2.2. Risperidone

A significant main effect of drug, F(3,16) = 40.84, P < .001, demonstrated that risperidone reduced motility of rats (P's < .001) on all doses tested (Fig. 4).

3.3. D-Amphetamine-disrupted PPI

3.3.1. SB-258741

As shown in Fig. 5A, there was a significant PPI effect, as reflected in the main effect of prepulse, F(2,66)=43.19, P < .001. The disruptive effect of D-amphetamine, F(1,33)=6.15, P < .02, was not reversed by SB-258741 as neither the main effect of drug, F(2,33)=0.08, P > .92, nor any of the interactions reached the significant level (*P*'s > .18). As shown in Fig. 5B, startle amplitude was not affected by the factors analysed (all *P*'s > .31).



Fig. 3. Effects of acute treatment with SB-258741 on spontaneous motility in rats. n=4 per drug group, but n=16 in VEH. *P < .05 versus VEH; ***P < .001 versus VEH.



Fig. 4. Effects of acute treatment with risperidone on spontaneous motility in rats. n=4 per drug group, but n=8 in VEH. ***P<.001 versus VEH.

3.3.2. Risperidone

As shown in Fig. 6A, there was a significant PPI effect, as reflected in the main effect of prepulse, F(2,84) = 102.63, P < .001. Risperidone normalised D-amphetamine-disrupted



Fig. 5. (A) Effects of acute treatment with SB-258741 on D-amphetaminedisrupted (2 mg/kg) PPI in rats. (B) Effects of SB-258741 on startle amplitude. n=6 in the VEH-AMPH, 2.3 mg/kg VEH, and 9.1 mg/kg groups, and n=7 in all the other groups.



Fig. 6. (A) Effects of acute treatment with risperidone on D-amphetaminedisrupted (2 mg/kg) PPI in rats. (B) Effects of risperidone on startle amplitude. n=8 per Drug × Treatment group. ***P<.001. ##P<.01 versus VEH-AMPH; ###P<.001 versus VEH-AMPH.

PPI, as reflected by the significant Treatment × Drug interaction, F(2,42) = 6.98, P < .003. Post hoc analysis indicated a significant difference only between vehicle and D-amphetamine groups in those animals receiving no treatment with risperidone (P < .001). Moreover, groups treated with risperidone at both doses in addition to D-amphetamine were significantly different from the AMPH–VEH-treated group (P's < .01). As shown in Fig. 6B, startle amplitude was not affected by the factors analysed (P's > .25).

3.4. PCP-disrupted PPI

3.4.1. SB-258741

As shown in Fig. 7A, there was a significant PPI effect, as reflected in the main effect of prepulse, F(2,84) = 98.61, P < .001. There was a significant disruptive effect of PCP reflected by the main effect of treatment, F(1,42) = 17.52, P < .001. SB-258741 enhanced PPI, as reflected by a significant effect of drug, F(2,42) = 6.49, P < .005. According to the double significant effect of treatment and drug, a Fisher PLSD post hoc test showed that there was a significant

difference between vehicle and PCP groups in the animal groups not treated with SB-258741 (P < .02) or treated with 2.3 mg/kg SB-258741 (P < .001). The animal group treated with SB-258741 at 9.1 mg/kg in addition to PCP was significantly different from the VEH–PCP-treated group (P < .003). As shown in Fig. 7B, a significant Treatment × Drug interaction, F(2,42)=3.93, P < .03, completed with a Fisher PLSD post hoc test confirmed that both doses of SB-258741 (P < .03) antagonised PCP-reduced startle amplitude (P=.053) in this experiment. Consequently, we cannot exclude the possibility that the effect of SB-258741 on the PCP-disrupted PPI model is simply a consequence of SB-258741 reversing PCP-reduced startle amplitude of rats.

3.4.2. Risperidone

As shown in Fig. 8A, there was a significant PPI effect, as reflected in the main effect of prepulse, F(2,84) = 66.96, P < .001. There was a significant disruptive effect of PCP, F(1,42) = 51.29, P < .001, which was dose-dependently reversed by risperidone as reflected by the significant Treatment × Drug interaction, F(2,42) = 3.80, P < .04. Post



Fig. 7. (A) Effects of acute treatment with SB-258741 on PCP (2 mg/kg)disrupted PPI in rats. (B) Effects of SB-258741 on startle amplitude. n=8per Drug × Treatment group. *P<.05; ***P<.001. #P<.05 versus VEH–PCP; ##P<.01 versus VEH–PCP.





Fig. 8. (A) Effects of acute treatment with risperidone on PCP (2 mg/kg)disrupted PPI in rats. (B) Effects of risperidone on startle amplitude. n=8per Drug × Treatment group. **P<.01; ***P<.001. #P<.05 versus VEH–PCP; ##P<.01 versus VEH–PCP.

hoc analysis indicated a significant difference between vehicle and PCP groups in the three treatment groups (P's < .003). However, this analysis showed that those groups treated with risperidone at both doses in addition to PCP were significantly different from the PCP–VEH-treated group (P's < .04). As shown in Fig. 8B, startle amplitude was not affected by factors analysed (all P's > .30).

3.5. PCP-disrupted SIT

3.5.1. SB-258741

3.5.1.1. Hyperactivity. A significant $Drug \times Treatment$ interaction, F(2,66) = 8.66, P < .001, showed that the various doses of SB-258741 interfered differently with PCP on activity (Fig. 9A). At the lowest dose (2.3 mg/kg), SB-258741 decreased activity similarly in the vehicle- and PCP-treated groups. At the highest dose (9.1 mg/kg), reduction of spontaneous activity induced by SB-258741 in addi-

tion to vehicle was much more pronounced than in the PCP-treated group.

3.5.1.2. Active SIT. As shown in Fig. 9B, PCP disrupted active SIT of rats, F(1,66)=88.14, P<.001, and SB-258741 reduced the level of active SIT of both vehicle-



Fig. 9. (A) Effects of subchronic (3 days) treatment with SB-258741 on PCP (2 mg/kg)-induced hyperactivity in rats. (B) Effects of subchronic (3 days) treatment with SB-258741 on PCP-disrupted SIT in rats. n=6 pairs of rats per Drug × Treatment group. Symbols: (\odot) SB-258741 in combination with vehicle; (\bigtriangledown) SB-258741 in combination with PCP. Open symbols indicate active SIT, and dark symbols indicate passive SIT **P*<.05 versus the respective vehicle-treated group. #*P*<.05 versus VEH–VEH or VEH–PCP.



Fig. 10. (A) Effects of subchronic (3 days) treatment with risperidone on PCP (2 mg/kg)-induced hyperactivity in rats. (B) Effects of subchronic (3 days) treatment with risperidone on PCP-disrupted SIT in rats. n=6 pairs of rats per Drug × Treatment group. Symbols: (\bigcirc) risperidone in combination with vehicle; (\bigcirc) risperidone in combination with PCP. Open symbols indicate active SIT, and dark symbols indicate passive SIT **P*<.05 versus the respective vehicle-treated group. #*P*<.05 versus VEH–VEH or VEH–PCP.

and PCP-treated rats, F(2,66) = 40.68, P < .001. A significant Drug × Treatment interaction, F(2,66) = 20.46, P < .001, as shown by a post hoc analysis demonstrated that 2.3 mg/kg of SB-258741 reduced the level of active SIT to the same extent in the vehicle- and PCP-treated groups. At 9.1 mg/kg, SB-258741 strongly reduced the level of active SIT in the vehicle-treated group in comparison to the PCP-treated group. This reduction of active SIT appears directly correlated to the reduction of activity described previously.

3.5.1.3. Passive SIT. As shown in Fig. 9B, SB-258741 given at 9.1 mg/kg induced marginal passive SIT, F(2,66) = 3.80, P < .05, which is also related to the reduction of spontaneous activity previously observed at this dose.

3.5.2. Risperidone

3.5.2.1. Hyperactivity. As shown in Fig. 10A, PCP induced hyperactivity in rats, F(1,66)=22.48, P<.001, and risperidone reduced the activity of these rats, F(2,66)=80.04, P<.001. A significant Drug × Treatment interaction, F(2,66)=8.66, P<.001, completed by a post hoc analysis showed that the lowest dose (0.16 mg/kg) of risperidone normalised hyperactivity in the PCP-treated groups and slightly reduced the spontaneous activity of rats. However, there was still a significant difference between the activity level of the VEH- and PCP-treated groups at this dose of risperidone. At the highest dose (0.31 mg/kg), the risperidone-induced decrease in activity was much more pronounced in the PCP-treated group than in the vehicle-treated group, with the consequence that both groups reached the same low level of activity.

3.5.2.2. Active SIT. As shown in Fig. 10B, PCP disrupted active SIT of rats, F(1,66) = 303.33, P < .001, and risperidone increased the level of active SIT of rats as demonstrated by a main effect of drug, F(2,66) = 3.99, P < .05. A significant Drug × Treatment interaction, F(2,66) = 6.73, P < .005, completed by a post hoc analysis demonstrated that at 0.16 mg/kg, risperidone did not reverse PCP-disrupted SIT. At 0.31 mg/kg, risperidone partially reversed this disruptive effect of PCP, without significant modification of the spontaneous SIT of rats.

3.5.2.3. Passive SIT. As shown in Fig. 10B, 0.31 mg/kg induced passive SIT, F(2,66) = 11.29, P < .001, which is correlated to the increase in active SIT previously described at this dose, and probably to the reduction of activity as well.



Fig. 11. Cataleptogenic effect of SB-258741 (dash line) and risperidone (full line). n=4 per drug group.

3.6. Catalepsy

3.6.1. SB-258741

As shown in Fig. 11, SB-258741 did not induce any catalepsy for doses up to 18 mg/kg and up to 24 h after administration (P's = 1).

3.6.2. Risperidone

As shown in Fig. 11, risperidone induced dose-dependent catalepsy, F(2,9) = 6.84, P < .02, for the two highest doses.

4. Discussion

This is the first attempt to study the potential antipsychotic activity of a selective 5-HT₇ receptor antagonist. SB-258741 is 200-fold selective over other 5-HT or dopaminergic receptors (see compound "13" in Lovell et al., 2000) and has significant effects in two models known to be predictive of antipsychotic action: D-amphetamineinduced hyperactivity in rats (Arnt, 1995, 2000) and PCPdisrupted PPI (Mansbach and Geyer, 1989; Bakshi and Geyer, 1995; Bakshi et al., 1994; Yamada et al., 1999). However, in both cases, this effect might be a consequence of a modification of motricity induced by SB-258741. SB-258741 dose-dependently antagonised D-amphetamine-induced hyperactivity but also reduced spontaneous activity of rats at the same doses (4.6 and 9.1 mg/kg). It is only at 2.3 mg/kg that SB-258741 antagonised specifically D-amphetamine-induced hyperactivity. It is also noted that rats treated with SB-258741 at this mid-dose were more active than VEH-AMPH rats when the effect of D-amphetamine elapsed. Thus, SB-258741 seems to have a dual effect on activity of rats. At high doses, it reduced activity of rats during the exploratory part of the test, but at lower doses, it keeps rats more active than VEH-AMPH-treated rats. This observation is difficult to explain, as we cannot conclude whether this effect is specific to SB-258741 or to a possible metabolite, due to a real pharmacodynamic effect of SB-258741 or to an effect on the pharmacokinetics of *D*-amphetamine. In comparison, risperidone dose-dependently reversed D-amphetamineinduced hyperactivity without inducing such hyperactivity at the end of the session. At the two highest doses risperidone also reduced activity of rats during the exploratory period, which questions the validity of the results obtained versus D-amphetamine in this model. In a similar fashion to SB-258741, risperidone significantly reduced the spontaneous activity of rats in the motility test when tested with doses that were effective in the D-amphetamine-induced hyperactivity model. Consequently, the effect of SB-258741 in this model is inconclusive as this compound antagonised D-amphetamine at doses that were similar to those inhibiting spontaneous activity of rats. However, risperidone, an antipsychotic with demonstrated clinical effect, suppresses locomotor activity at the same doses that were effective in the D-amphetamine-induced hyperactivity model.

SB-258741 also dose-dependently normalised the effect of PCP in PCP-disrupted PPI in rats. However, on both doses tested, SB-258741 also normalised the reduction of startle amplitude induced by PCP. Thus, the beneficial effect of SB-258741 in PCP-disrupted PPI is certainly not independent of its effect on the startle amplitude of rats in the PCP-treated groups. This means that the positive effect of SB-258741 in this model is probably not related to a sensory gating process (Swerdlow et al., 2000). Consequently, the beneficial effect of SB-258741 in the PCP-disrupted PPI model should be considered carefully, and it seems difficult to predict an antipsychotic action for SB-258741 in this model as well. Contrary to SB-258741, risperidone antagonised PCP-disrupted PPI without effect on startle amplitude and under condition at which PCP did not affect startle amplitude as well. This result with risperidone confirms another previous report (Yamada et al., 1999) but is contrary to another publication showing that risperidone was not effective in this model (Swerdlow et al., 1996). It shows that like clozapine (Bakshi et al., 1994; Yamada et al., 1999; Johansson et al., 1994), the antipsychotic-like effect of risperidone in this model is variable between laboratories.

On D-amphetamine-disrupted PPI, a model predictive of antipsychotic action (Swerdlow and Geyer, 1993; Paabøl Andersen and Pouzet, 2001), SB-258741 did not reverse the effect of D-amphetamine. This effect is opposite to the one obtained with risperidone but is consistent with the lack of clear effect obtained with SB-258741 on both the D-amphetamine-induced hyperactivity and PCPdisrupted PPI models. Thus, according to results obtained in these three models, the probability that SB-258741 can, on its own, treat positive symptoms in schizophrenic patients seems weak.

With respect to the model indicative of effect on negative symptoms of schizophrenia, i.e., PCP-disrupted SIT in rats (Sams-Dodd, 1997, 1998), SB-258741 did not show any beneficial effect, as it enhanced the disruptive effect of PCP on active SIT instead of reducing it. Contrary to the motility test, all doses tested for both SB-258741 and risperidone reduced spontaneous activity. This difference can certainly be explained by different experimental conditions (acute versus 3 days of treatment; macrolon type III cages versus large arena as boxes for testing). However, only risperidone antagonised PCP-disrupted SIT, although passive SIT was increased to the same extent. Consequently, specific antagonism of the 5-HT₇ receptor does not seem to explain the beneficial effect of risperidone observed in this study and previously reported for this model (Sams-Dodd, 1997). However, the possibility that a combination of the effect of 5-HT₇ receptor antagonism plus the dopaminergic D_2 and/or 5-HT_{2A} receptor antagonism of risperidone has a positive effect in this model cannot be excluded.

As regards side effects, SB-258741 tested at 18 mg/kg did not induce catalepsy. As this model is a predictor of

EPS (Arnt, 1982), we can exclude the possibility that antagonism on the 5-HT₇ receptor would induce EPS. This lack of cataleptogenic effect of SB-258741 is in contrast to the induction of catalepsy obtained with risperidone in this study. As previously suggested, we cannot exclude the possibility that a combination of antagonism on the 5-HT₇ receptor plus the dopaminergic D₂ and/or 5-HT_{2A} receptor would change the pattern of results obtained both in models predictive of antipsychotic action and EPS. However, a preliminary study conducted in our laboratory, testing SB-258741 in combination with the dopaminergic D_2 receptor antagonist remoxipride, or the 5-HT_{2A} receptor antagonist MDL-100151 in another model predictive of antipsychotic action, PCP-induced hyperactivity in mice (Gleason and Shannon, 1997), did not show any enhancement of the beneficial effect of these two compounds (unpublished data).

This study demonstrated that we should not expect to obtain an antipsychotic action with SB-258741. According to the specificity of this compound (see compound "13" in Lovell et al., 2000), it seems doubtful that other specific 5-HT₇ receptor antagonists would have any antipsychotic action on their own as well. On the other hand, a lack of information about the pharmacokinetics of this compound does not permit exclusion of the possibility that its central effect is weak due to a poor brain penetrability. However, two arguments go against this hypothesis. First, we described in this study that SB-258741 antagonised the effect of PCP on startle amplitude of rats but did not affect the spontaneous startle amplitude on its own. This suggests an effect of this compound on central structures involved in regulation of startle. Second, SB-258741 and SB-269970 have very similar chemical structures (Lovell et al., 2000), with similar biological activities, and SB-269970 is known to have a good brain penetrability (Hagan et al., 2000).

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References

- Arnt J. Pharmacological specificity of conditioned avoidance response inhibition in rats: inhibition by neuroleptics and correlation to dopamine receptor blockade. Acta Pharmacol Toxicol 1982;51:321–9.
- Arnt J. Differential effects of classical and newer antipsychotics on the hypermotility induced by two dose levels of D-amphetamine. Eur J Pharmacol 1995;283:55–62.
- Arnt J. Screening models for antipsychotic drugs. In: Ellenbroek BA,

Cools AR, editors. Atypical antipsychotics. Basel: Bikhauser, 2000. pp. 99-119.

- Arnt J, Skarsfeldt T. Do novel antipsychotics have similar pharmacological characteristics? A review of the evidence. Neuropsychopharmacology 1998;18(2):63–101.
- Bakshi VP, Geyer MA. Antagonism of phencyclidine-induced deficits in prepulse inhibition by the putative atypical antipsychotic olanzapine. Psychopharmacology 1995;122:198–201.
- Bakshi VP, Swerdlow NR, Geyer MA. Clozapine antagonizes phencyclidine-induced deficits in sensorimotor gating of the startle response. J Pharmacol Exp Ther 1994;271:787–94.
- Blind O. A comparative review of new antipsychotics. Can J Psychiatry 1999;44:235-44.
- Braff DL, Geyer MA. Sensorimotor gating and the neurobiology of schizophrenia: human and animal model studies. Arch Gen Psychiatry 1990; 47:181–8.
- Braff DL, Grillon C. Gating and habituation of the startle reflex in schizophrenic patients. Arch Gen Psychiatry 1992;49:206–15.
- Branchek TA, Gustafson MM, Durkin MM, Bard JA, Weinshank RL. Autoradiographic localization of 5-ht₇ and its mRNA in rats CNS by radioligand binding and in situ hybridization histochemistry. Br J Pharmacol 1995;112:110P.
- Corbett R, Camacho F, Woods AT, Kerman LL, Fishkin RJ, Brooks K, Dunn RW. Antipsychotic agents antagonize non-competitive *N*-methyl-D-aspartate antagonist-induced behaviors. Psychopharmacology 1995; 120:67–74.
- Davis M. Apomorphine, D-amphetamine, strychnine and yohimbine do not alter prepulse inhibition of the acoustic startle reflex. Psychopharmacology 1988;95:151–6.
- Doyland PS, Eastwood S, Ellis C, Bergsma D, Jones RJ, Cloger IS, Upton N, Middlemiss DN. High specific activity [³H]5-CT binding: correlation of guinea-pig cortex with human cloned 5-ht₇ receptors. Proc Br Pharmacol Soc. 1995;3.
- Druhan JP, Geyer MA, Valentino RJ. Lack of sensitization to the effects of D-amphetamine and apomorphine on sensorimotor gating in rats. Psychopharmacology 1998;135:296–304.
- Eglen RM, Jasper JR, Chang DJ, Martin GR. The 5-HT₇ receptor: orphan found. Trends Pharmacol Sci 1997;18:104–7.
- Forbes IT, Dabbs S, Duckworth DM, Jennings AJ, King FD, Lovell PJ, Brown AM, Collin L, Hagan JJ, Middlemiss DN, Riley GJ, Thomas DR, Upton N. (*R*)-3,*N*-Dimethyl-*N*-[1-methyl-3-(4-methyl-piperidin-1-yl)propyl]benzenesulfonamide: the first selective 5-HT₇ receptor antagonist. J Med Chem 1998;41:655–7.
- Glazer WM. Extrapyramidal side effects, tardive dyskinesia, and the concept of atypicality. J Clin Psychiatry 2000;61:16–21.
- Gleason SD, Shannon HE. Blockade of phencyclidine-induced hyperlocomotion by olanzapine, clozapine, and serotonin receptor subtype selective antagonists in mice. Psychopharmacology 1997;129:79–84.
- Goldstein JM. The new generation of antipsychotic drugs: how atypical are they? Int J Neuropsychopharmacol 2000;3:339–49.
- Hagan JJ, Price GW, Jeffrey P, Deeks NJ, Stean T, Piper D, Smith MI, Upton N, Medhurst AD, Middlemiss DN, Riley GJ, Lovell PJ, Bromidge SM, Thomas DR. Characterization of SB-269970-A, a selective 5-HT₇ receptor antagonist. Br J Pharmacol 2000;130:539–48.
- Johansson C, Jackson DM, Svensson L. The atypical antipsychotic, remoxipride, blocks phencyclidine-induced disruption of prepulse inhibition in the rat. Psychopharmacology 1994;116:437–42.
- Johansson C, Jackson DM, Zhang J, Svensson L. Prepulse inhibition of acoustic startle, a measure of sensorimotor gating: effects of antipsychotics and other agents in rats. Pharmacol, Biochem Behav 1995;52: 649–54.
- Le Corre S, Sharp T, Young AH, Harrison PJ. Increase of 5-HT₇ (serotonin-7) and 5-HT_{1A} (serotonin-1A) receptor mRNA expression in rat hippocampus after adrenalectomy. Psychopharmacology 1997;130:368–74.
- Lena Mullins U, Gianutsos G, Eison AS. Effects of antidepressants on 5-HT₇ receptor regulation in the rat hypothalamus. Neuropsychopharmacology 1999;21:352–67.

- Lovell PJ, Bromidge SM, Dabbs S, Duckworth DM, Forbes IT, Jennings AJ, King FD, Middlemiss DN, Rahman SK, Saunders DV, Collin LL, Hagan JJ, Riley GJ, Thomas DR. A novel, potent, and selective 5-HT₇ antagonist: (*R*)-3-(2-(2-(4-methylpiperidin-1-yl)-ethyl)pyrrolidine-1-sulfonyl)phenol (SB-269970). J Med Chem 2000;43:342–5.
- Lovenberg TW, Baron BM, Lecea L, Miller JD, Prosser RA, Rea MA, Foye PE, Racke M, Slone AL, Siegel BW, Danielson PE, Sutcliffe JG, Erlander MG. A novel adenylyl cyclase-activating serotonin receptor (5-HT₇) implicated in the regulation of mammalian circadian rhythms. Neuron 1993;11:449–58.
- Mansbach RS, Geyer MA. Effects of phencyclidine and phencyclidine biologs on sensorimotor gating in the rat. Neuropsychopharmacology 1989;2(4):299–308.
- Mansbach RS, Geyer MA, Braff DL. Dopaminergic stimulation disrupts sensorimotor gating in the rat. Psychopharmacology 1988;94: 507–14.
- Meneses A, Terrón JA. Role of 5-HT1A and 5-HT7 receptors in the facilitatory response induced by 8-OH-DPAT on learning consolidation. Behav Brain Res 2001;21:21–8.
- Paabøl Andersen M, Pouzet B. Effects of acute versus chronic treatment with typical or atypical antipsychotics on D-amphetamine-induced sensorimotor gating deficits in rats. Psychopharmacology 2001;156: 291–304.
- Roth BL, Craigo SC, Salman Choudhary M, Uluer A, Monsma FJ, Shen Y, Meltzer HY, Sibley DR. Binding of typical and atypical antipsychotic agents to 5-hydroxytryptamine-6 and 5-hydroxytriptamine-7 receptors. J Pharmacol Exp Ther 1994;268(3):1403–10.
- Sams-Dodd F. Automation of the social interaction test by a video-tracking system: behavioural effects of repeated phencyclidine treatment. J Neurosci Methods 1995;59:157–67.
- Sams-Dodd F. Phencyclidine-induced stereotyped behaviour and social isolation in rats: a possible animal model of schizophrenia. Behav Pharmacol 1996;7:3–23.
- Sams-Dodd F. Effect of novel antipsychotic drugs on phencyclidine-induced strereotyped behaviour and social isolation in the rat social interaction test. Behav Pharmacol 1997;8:196–215.
- Sams-Dodd F. A test of the predictive validity of animal models of schizophrenia based on phencyclidine and D-apomorphine. Neurophychopharmacology 1998;18(4):293–304.
- Schwartz WJ. A clinician's primer on the circadian clock: its localization, function and resetting. Adv Intern Med 1993;38:81–106.

Sleight AJ, Carolo C, Petit N, Zwingelstein C, Bourson A. Identification of

5-HT₇ receptor binding sites in rat hypothalamus. Mol Pharmacol 1995; 47:99-103.

- Swerdlow NR, Geyer MA. Clozapine and haloperidol in an animal model of sensorimotor gating deficits in schizophrenia. Pharmacol, Biochem Behav 1993;44:741–4.
- Swerdlow NR, Braff DL, Geyer MA, Koob GF. Central dopamine hyperactivity in rats mimics abnormal acoustic startle response in schizophrenics. Biol Psychiatry 1986;21:23–33.
- Swerdlow NR, Braff DL, Taaid N, Geyer MA. Assessing the validity of an animal model of deficient sensorimotor gating in schizophrenic patients. Arch Gen Psychiatry 1994;51:139–54.
- Swerdlow NR, Bakshi VP, Geyer MA. Seroquel restores sensorimotor gating in phencyclidine-treated rats. J Pharmacol Exp Ther 1996;279(3): 1290–9.
- Swerdlow NR, Braff DL, Geyer MA. Animal models of deficient sensorimotor gating: what we know, what we think we know, and what we hope to know soon. Behav Pharmacol 2000;11(3–4):185–204.
- Terrón JA, Falcón-Neri A. Pharmacological evidence for the 5-HT₇ receptor mediating smooth muscle relaxation in canine cerebral arteries. Br J Pharmacol 1999;127:609–16.
- Thomas DR, Middlemiss DN, Taylor SG, Nelson P, Brown AM. 5-CT stimulation of adenyl cyclase activity in guinea-pig hippocampus: evidence for involvement of 5-HT₇ and 5-HT_{1A} receptors. Br J Pharmacol 1999;128:158–64.
- To ZP, Bonhaus DW, Eglen RM, Jakeman LB. Characterization and distribution of putative 5-HT₇ receptors in the guinea-pig brain. Br J Pharmacol 1995;115:107–16.
- Tsou A, Kosaka A, Bach C, Zuppan P, Yee C, Tom L, Alvarez R, Ramsey S, Bonhaus DW, Stefanich E, Jakeman L, Eglen RM, Chan HW. Cloning and expression of a 5-HT₇ receptor positively coupled to adenylyl cyclase. J Neurochem 1994;63:456–64.
- Yamada S, Harano M, Annoh N, Nakamura K, Tanaka M. Involvement of serotonin 2A receptors in phencyclidine-induced disruption of prepulse inhibition of the acoustic startle in rats. Biol Psychiatry 1999;46:832–8.
- Yau JLW, Noble J, Widdowson J, Seckl JR. Impact of adrenalectomy on the 5-HT₆ and 5-HT₇ receptor gene expression in the rat hippocampus. Mol Brain Res 1997;45:182–6.
- Zhang J, Forkstam C, Engel JA, Svensson L. Role of dopamine in prepulse inhibition of acoustic startle. Psychopharmacology 2000;149:181–8.
- Zhukovskaya NL, Neumaier JF. Clozapine downregulates 5-hydroxytryptamine₆ (5-HT₆) and upregulates 5-HT₇ receptors in Hela cells. Neurosci Lett 2000;288:236–40.